



Chromosomal variations and cytotaxonomical considerations in two populations of *Nitella hyalina* (Charophyceae, Characeae) from West Bengal, India

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Abstract

Chromosome number and morphological variation of two populations of *N. hyalina* (DC.) Ag. have been studied. Chromosome morphology was worked out from well spreadout metaphase plates from dividing antheridial filament cells. Population I collected from the bed of Ajoy river (district Burdwan) shows $n=18$ chromosomes. Population II collected from the bed of the river Kopai (district Birbhum) shows $n=21$ chromosomes. The karyotype formula for population I is $L (Sm_7+m_1+St_0) + M (Sm_4+m_2+St_0) + S (Sm_2+m_2+St_0)$. The karyotype formula for population II is $L (Sm_1 + m_2+ St_0) + M (Sm_{12}+ m_3+ St_0) + S (Sm_1+m_2+St_0)$. Total form percent (TF%) calculated for population I and II are 38.21 and 38.25 respectively. These two populations do not show any variation in morphological features but variation in chromosome number and morphology is evident. Occurrence of different ploidy level and symmetrical nature of karyotype for *N. hyalina*, as compared to other species of *Nitella*, is established.

Key words: Charophyceae, Characeae, Chromosome, Cytotaxonomy, *Nitella hyalina*, West Bengal.

Introduction

Cytological data with reference to chromosome number, morphology and karyotype pattern provide much information in tracing origin, affinities and interrelationship of plants including algae (Ehrendorfer *et al.*, 1968; Sharma, 1969; Stebbins 1971). Investigation on higher plants with regard to karyotype and karyogram has helped in establishing evolutionary status for a taxon and relationship between different taxonomic groups. However, any definite systematic conclusion in algae on the basis of chromosome data is very difficult because it is not easy to reveal the exact chromosome morphology.

In the members of the order Charales centromeric position and chromosome size variation could be studied with perfection because of excellent metaphase chromosome preparations achieved by the application of proper pretreatment and staining schedule (Dyer, 1963; Sawa, 1965; Ray and Chatterjee, 1989; Ray, 1998). Kanahori (1974) refined techniques for examination of chromosomes from vegetative shoot apices of Charophytes in a study of *Nitella hyalina*. To get a clear picture of chromosome profile from karyotype of different species four different criteria are taken into account viz. (i) variation in absolute chromosome size, (ii) variation in the position of centromere, (iii) variation in relative chromosome size, (iv) variation in basic number. Morphological characters primarily considered to describe and distinguish various taxa, under Charales, are simultaneously considered with cytological data to draw a clear demarcating line between different species and intraspecific taxa. This approach helps to ascertain the validity of the evolutionary status of various taxa, established on the basis of morphological characters only (Chatterjee, 1976 a, b, 1979 a, b; Ray, 1998; Ray and Chatterjee 1988a, b, 1989, 1992, 1994; Mandal and Ray, 2000). Chakrabarty *et al.* (2009) has established definite correlation through CCA ordination study in *Chara fibrosa* complex between morphological features and chromosome number.

Cytology and cytotaxonomy of different species of *Nitella* have extensively been studied from different parts of India (Sarma and Khan, 1964, 1965; Khan and Sarma, 1967; Ramjee and Sarma, 1971; Ramjee and Bhatnagar, 1978 a, b; Subramanyam and Chowdhury, 1992; Pundhir *et al.*, 1993). All these studies were based on cytological data obtained from unpretreated materials. A large number of species and interspecific taxa belonging to *Nitella* have been studied from West Bengal and significant cytotaxonomic conclusions have been drawn (Ray and Chatterjee, 1983, 1988b, 1994; Ray, 1998; Mandal and Ray, 2001). Using the refined technique (combination of pretreatment and lacto-propiono orcein staining) the chromosomes were worked out both from vegetative apices as well as

antheridial filament cells. Ray and Chatterjee (1983) published the chromosome number and Karyogram of *N. acuminata* var. *acuminata* f. *acuminata* A. BR. Ex. WALLM. New chromosome number (n=18) and karyogram, worked out from vegetative cells, of *N. stuartii* A. BR. was reported from West Bengal (Ray and Chatterjee, 1988b). All these cytological works have been worked out with technique involving pretreatment and Lacto-propiono orcein staining (Ray and Chatterjee, 1989). Cytotaxonomic work based on karyotype analysis at the subspecific level in *N. furcata* complex was done for the first time by Ray and Chatterjee (1994). All these works involved correlation of morphological and cytological variations and the significance of chromosome data in characterization of the taxa of *Nitella*. Mandal and Ray (2001) reported for the first time the cytology of *N. hyalina* var. *hyalina* f. *hyalina* and f. *formosa* (T. F. A.) R.D.W. from West Bengal having chromosome number n=18 which is similar to earlier reports from India and abroad (Guerlesquin, 1963; Hotchkiss, 1965; Sarma and Khan, 1965; Kanahori, 1974; Peshkov *et al.*, 1974). Considering the variations found in cytological profile of various taxa under *N. hyalina* as reported by earlier workers, a further study of *N. hyalina* populations has been undertaken in this investigation.

Material and methods

(a) *Geographical location*: Populations *N. hyalina* were collected from the districts of Burdwan and Birbhum of the State of West Bengal having similar weather condition. Both the locations of collection have Vindiyian alluvial type of soil. Within each location four permanent water bodies with charophyte population were identified.

(b) *Morphological identification*: The species was identified following Wood (1965).

(c) *Cytological Methods*: Combination of Lacto-propiono orcein staining and pretreatment as established by Ray and Chatterjee (1989) has been applied here. Metaphase stage of mitotically dividing cells of antheridial filament were considered for karyotype study in most of the cases. The young shoot tips were collected from the natural aquatic habitats or from soil-water culture established in the net house. The tip portions were observed under dissecting microscope for detection of correct stage of antheridium for cytological study.

i) *Pretreatment*: Thoroughly washed tip portions were pretreated in a chilled mixture of 0.1% aqueous solution of Colchicine and 0.002M aqueous solution of 8-hydroxyquinoline. Then the apical portion with antheridia was put into the chilled pretreatment chemical. The antheridia were allowed to stay there at 16°C-18°C for 2-2½ hrs.

ii) *Fixation*: After pretreatment the material was fixed in freshly prepared 1:3 mixture of Propionic acid and Absolute alcohol for 2 hours.

iv) *Treatment with 0.1N HCl*: After fixation, the shoot tip along with the antheridia was softened in 0.1N HCl at 60°C for 3-4 minutes.

v) *Washing in water and 45% propionic acid*: The material was washed in distilled water to remove the HCl. The water was decanted and the material was washed with 45% Propionic acid and kept in the propionic acid for 3-5 minutes.

vi) *Staining*: The material was stained in 2% Lacto-propiono Orcein (Dyer, 1963) overnight.

vii) *Squashing*: After staining, the material was put into 45% propionic acid. The antheridial filaments were teased out from the antheridia under a binocular dissecting stereomicroscope and the final squashing of the antheridial filaments were done in 45% propionic acid under a cover slip.

Chromosome morphology was designated according to Levan *et al.* (1964) on the basis of centromeric index value. Total form percent (TF %) was calculated for each taxon using the formula suggested by Huziwara (1962). Chromosomes were further categorized according to their length (Khan and Sarma, 1967a).

Results

Morphological features

Plants monoecious; upper whorls crowded and covered with mucus; internodes 2 – 4 times as long as branchlets; branchlets heteroclemous; fertile branchlets 8 in a whorl; 2 – 3 furcate; sterile similar to fertile; dactyls 2-celled, 4 – 6; basal cell tapering, end cell conical, acute; gametangia conjoined at all branchlet furcations; oogonia solitary; oospore

reddish brown, striae thin, ridges prominent. Two populations resembled each other with regard to taxonomically significant morphological features.

Cytological features

Population - I of *N. hyalina* collected from Ajay river bed (District of Burdwan) shows $n=18$ chromosomes (Fig.1a, 2a). The chromosome length varies from 2.50 to 5.63 μm and the total chromosome length is 76.91 μm . There are 13 submedian (Sm), 5 median (m) chromosomes (Fig.2b). The Karyotype formula is $L (Sm_7+m_1+St_0) + M (Sm_4+m_2+St_0) + S (Sm_2+m_2+St_0)$, where L, M and S indicate long, medium, and short chromosomes and zero represents the absence of a particular type of chromosome. The population - II from Kopai (District of Birbhum) is found to have $n=21$ chromosomes (Fig.1b, 2c) and the length of the chromosomes varies from 2.50 - 6.88 μm . The total length of the chromosome set was 80.07 μm . There are 14 Sm, 7 m chromosomes (Fig.2d). The karyotype formula constructed for this population is $L (Sm_1 + m_2 + St_0) + M (Sm_{12} + m_3 + St_0) + S (Sm_1+m_2+St_0)$. The TF% values were 38.21 and 38.25 for the population with $n=18$ and for the population with $n=21$ respectively.

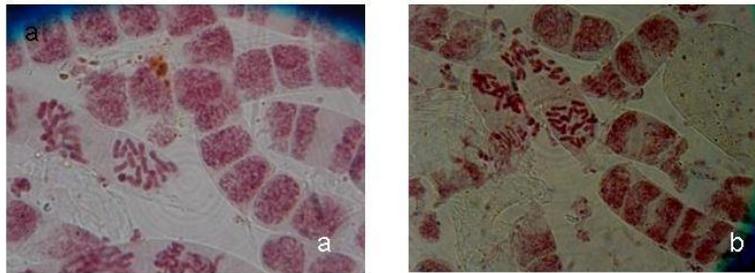


Fig. 1: Metaphase plate in antheridial filament cells of *N. hyalina* showing (a) $n = 18$ chromosomes in population I, (b) $n = 21$ chromosomes in population II.

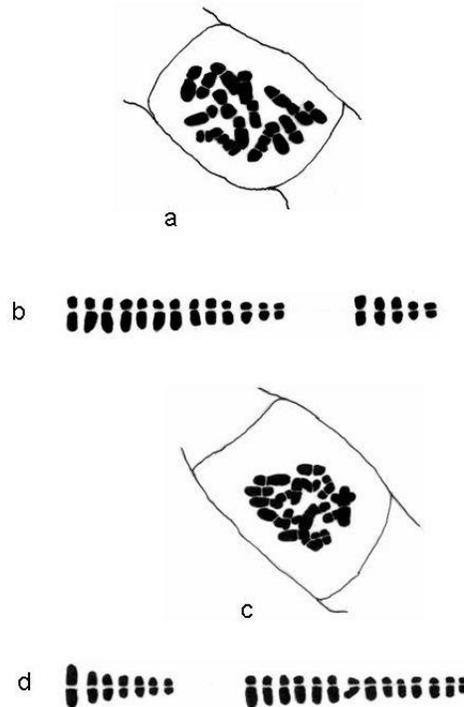


Fig. 2: Population I (a) karyotype, (b) karyogram showing 13 submedian and 5 median chromosomes. Population II (c) karyotype, (d) karyogram showing 14 submedian and 7 median chromosomes.

Table-1: Chromosome number of various populations of *N. hyalina* from West Bengal.

Sl. No.	Name of Taxon	Location	Chromosome No.	Reference
1	<i>Nitella hyalina</i>	Medinipur	n=18	Ray, 1988
2	<i>N. hyaline</i>	Birbhum	n=18	Mandal and Ray, 2001
3	<i>N. hyalina</i>	Birbhum	n = 18	Present author
4	<i>N. hyaline</i>	Birbhum	n = 21	Present author
5	<i>N. hyalina</i>	Burdwan	n = 18	Present author

Discussion

In addition to the morphological features (vegetative and reproductive), several other parameters have been taken into consideration for the identification of the species of *Nitella*. One of the most important parameters considered is the chromosome number, morphology and karyogram (Mandal and Ray, 2001). Ray and Chatterjee (1988b, 1992, 1994) clearly indicated the importance of chromosome number, morphology and karyotype in the classification of various interspecific taxa belonging to *C. fibrosa* species complex, *C. zeylanica* and *N. furcata*. Standardization of cytological technique by Ray and Chatterjee (1989) and adoption of staining technique of Dyer (1963) have helped to ascertain the chromosome number and morphology with precision in charalean species. As the various populations of *N. hyalina* were being studied with the help of revised cytological procedure two populations with different ploidy levels were found from this geographic region at two locations having similar soil type but separated by a distance of 10 km. Cosmopolitan taxa like *N. acuminata* and *N. hyalina* show wide ranging chromosome number n=9, 12, 15, 18, 21 (Ray, 2005).

Nitella hyalina belongs to section Decandollea (Wood, 1965). *Nitella hyalina* var. *hyalina* f. *hyalina* is a monoecious species with heteroclemous branching. This taxon was studied cytologically in India by Sarma and Khan (1964), Ramjee and Sarma (1971), Ramjee and Bhatnagar (1978), Ray (1998) and Chromosome number reported in every case is n= 18. A chromosome number of n=18 was found in representatives of this taxon in Sweden (Walther, 1929), Japan (Sato, 1959; Kanahori, 1974), France (Guerlesquin, 1963), New Caledonia, US, and Mexico (Hotchkiss, 1964). It is interesting to note that Kanahori (1974) studied two clones of *N. hyalina* (n=18) occurring in Japan and noted differences in chromosome length and arm ratio of the two clones studied from somatic cells.

In the present investigation, 2 populations of *N. hyalina* were studied. Owing to the clarity of the primary constriction, the categorization of the chromosome was possible. The TF% value of 38.21 (n=18) and 38.25 (n=21) for these two populations of *N. hyalina* var. *hyalina* f. *hyalina* reveals a relative symmetrical karyotype and thus is evidence of relatively primitive status of this heteroclemous taxon among the species of *Nitella*. It is pertinent to mention here that *N. stuartii* another heteroclemous taxon shows TF%=38.21 (Ray and Chatterjee, 1988b) while *N. acuminata* a homoclemous taxon show TF%= 32.2 (Ray and Chatterjee, 1983). Primitive nature of *N. hyalina* as compared to homoclemous taxa like *N. acuminata* is thus evident. The earlier findings that n=18 is a deep seated number and wide spread among the species of *Nitella* and that there is common occurrence of euploidy in this species complex is confirmed. Additionally, this part of India is added in the list of regions where populations of *N. hyalina* with varied ploidy level are found.

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